

Lightweight 3D Reconstruction of Large Fluorescence Microscopy Volumes via Statistical and Deep Learning Segmentation

1 THOMAS LUND¹ AND JAN MULDER²

2 ¹*Carleton College*

3 ²*Karolinska Institute*

4 (Dated: January 2026)

5 1. INTRODUCTION

6 Recent improvements in z stacked imaging methods
7 such as confocal laser scanning microscopy and light
8 sheet fluorescence microscopy have made large and high
9 quality datasets available for many biological objects of
10 interest. These datasets are crucial for various applica-
11 tions, including protein expression studies, blood vessel
12 mapping, and nerve signal tracing. Because of the com-
13 putational difficulty of rendering these datasets, their
14 study is often restricted to those with access to high per-
15 formance compute resources, in particular system mem-
16 ory, with typical rendering techniques often requiring in
17 the hundreds of gigabytes to hold the full models ef-
18 ficiently. In this study, we focus on a three channel,
19 volumetric 2056x2048x2022 dataset derived from a flu-
20 orescence microscopy scan of a mouse heart [Uhlén et al.](#)
21 ([2015](#)), where blood vessel walls and nerve fibers were
22 stained. We present a preprocessing and segmentation
23 pipeline that converts multi-channel fluorescence images
24 into voxelized binary masks corresponding to structures
25 of interest. Similar works have also attempted to seg-
26 ment z stacked imaging on smaller datasets through
27 varying segmentation methods. [Zekri & Lang \(2024\)](#)
28 Our approach enables efficient downstream analysis and
29 visualization of large-scale volumetric data on more
30 modest computational hardware. This strategy facili-
31 tates access to whole organ imaging datasets and sup-
32 ports further analysis of complex biological structures.

33 2. INSTRUMENTAL LIMITATIONS

34 Multiplex immunohistochemistry and immunofluores-
35 cence (mIHC/IF) are powerful techniques for visualizing
36 multiple protein markers within tissue sections, enabling
37 the detailed study of complex biological structures. In
38 this study mIHC/IF were employed to visualize protein
39 markers in tissue sections, with each image representing
40 the summed average of several frames. This approach
41 precluded the deconvolution of the exact point spread
42 function. Because each fluorescent channel was captured

43 separately, the resulting frames did not align perfectly in
44 the z-axis, further complicating the accuracy of spatial
45 registration. Additionally, variations in focus due to the
46 tissue's distance from the scanner caused a progressive
47 loss of sharpness toward the periphery of each frame.
48 A further complication arose from the heart wall, which
49 trapped fluorescent chemicals during staining, leading to
50 false-positive signals in the blood vessel scans. Finally,
51 the intensity of each image varied, and there is an un-
52 even illumination pattern within images, leading to in-
53 consistent bright and dark portions of each slice. These
54 factors necessitate significant pre-processing and require
55 a more robust approach than an intensity threshold.

56 3. DATA PIPELINE

57 To reduce the data footprint while maintaining the full
58 spatial accuracy, we first convert the multi-channel im-
59 ages into voxelized binary masks containing only voxels
60 which had the features of interest.

61 3.1. *Preprocessing*

62 We begin by normalizing the intensity values across
63 the z-direction of the full dataset one channel at a time
64 to reduce image to image intensity variance. This nor-
65 malization helps ensure consistency across slices of the
66 z-stack and mitigate any depth-dependent variations in
67 signal intensity. Next, we apply an inverse hyperbolic
68 sine (asinh) filter to stretch each slice, which effectively
69 preserves relative intensity levels before we compress the
70 dynamic range[fig.3]. We then apply a Contrast Lim-
71 ited Adaptive Histogram Equalization [Zuiderveld \(1994\)](#)
72 (CLAHE) to improve local contrast and flatten back-
73 ground intensities. CLAHE operates by enhancing the
74 contrast within small local regions of the image, which
75 scales each local intensity to a standard range across the
76 whole image, evening out the slice, and enhancing con-
77 trast between background and fluorescing features[fig.2].
78 To maximize the intensity flattening, we set the clipping
79 limit to 0.01 and used a window size of 32x32 pixels.

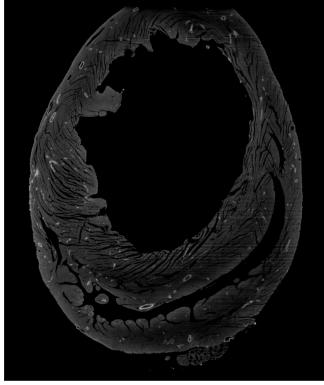


Figure 1. Scaled image slice before CLAHE

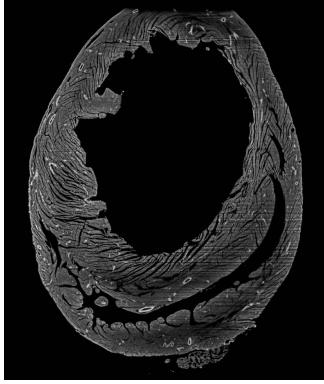


Figure 2. Scaled image slice after CLAHE

⁸⁰ Finally, we use a robust scaler to remove outliers and
⁸¹ normalize the data to the final 0-255 greyscale range.

⁸² 3.2. *U-Net*

⁸³ For image segmentation, we use a 9-layer U-Net architecture, which is well-suited for biomedical image segmentation tasks due to its ability to capture both local and global features Ronneberger et al. (2015). The ⁸⁷ model accepts input images of size 256x256 pixels. To ⁸⁸ prepare the original dataset, we tile the data slices into ⁸⁹ smaller 256x256 pixel tiles, which allows the model to ⁹⁰ handle high-resolution images efficiently. We manually ⁹¹ segmented a small set of 32 tiles to create ground truth ⁹² masks for training to test this model. This limited training ⁹³ set was used to evaluate the feasibility of the approach, a larger training dataset would likely increase ⁹⁵ performance. To supplement the size of the training ⁹⁶ set and improve model generalization, we applied standard ⁹⁷ data augmentation like rotation, translation and ⁹⁸ noise generation. For training, we used Tversky-Focal ⁹⁹ loss as our loss function. This loss function is designed ¹⁰⁰ to handle class imbalance, where the background often ¹⁰¹ dominates the foreground structures, like is the case for ¹⁰² potentially single-pixel nerve fibers or capillaries. To im-

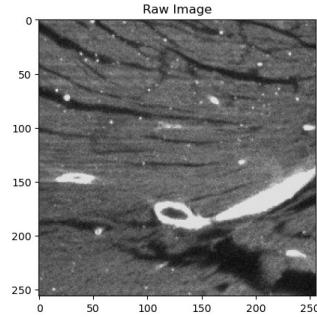


Figure 3. 256x256 input to U-Net model

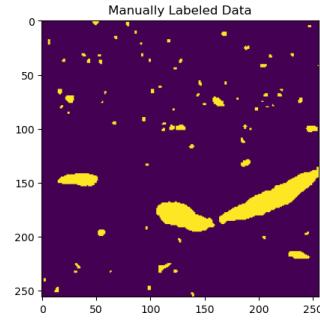


Figure 4. Manually labeled mask taken from the validation set

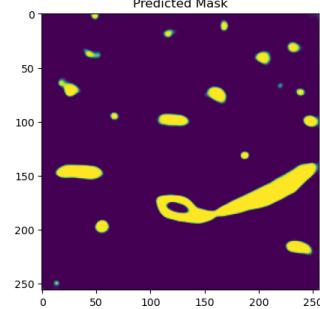


Figure 5. Predicted mask for the same image

¹⁰³ prove fitting performance we used a training loop with
¹⁰⁴ early stopping at loss plateauing, learning rate adap-
¹⁰⁵ tation for oscillating loss, and model checkpoints that
¹⁰⁶ save the best model based on loss values on the test set.
¹⁰⁷ Training was carried out on 50 epochs, and the early
¹⁰⁸ stopping filter did not trigger. We had a best fit at loss
¹⁰⁹ = 0.55, and training accuracy = 0.98. Despite achieving
¹¹⁰ high overall accuracy, small structures such as capillaries
¹¹¹ and nerve fibers were not segmented effectively by the
¹¹² model. To address this issue, we applied a difference of
¹¹³ median filter, which helped to enhance the segmentation
¹¹⁴ of these finer structures.

¹¹⁵ 3.3. *Difference of Medians*

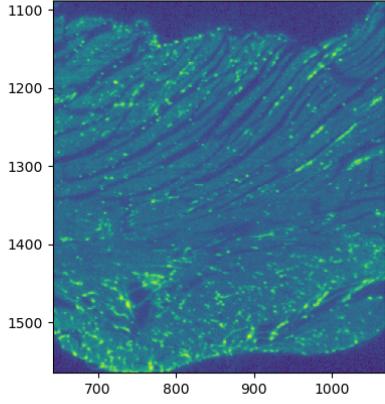


Figure 6. Zoomed section of a slice from the nerve fiber imaging

116 To enhance medium-scale fluorescent structures in the
 117 grayscale images, we apply a difference-of-medians fil-
 118 tering approach designed to suppress both large-scale
 119 background features and high-frequency noise. For each
 120 image slice, we compute two median-filtered versions:
 121 one using a large kernel[fig.8] (15×15 pixels) and a
 122 second using a small kernel[fig.7] (3×3 pixels). The
 123 large-kernel median filter captures slowly varying, large-
 124 scale intensity components, such as organ boundaries
 125 and broad illumination gradients, while effectively av-
 126 eraging out smaller structures of interest. This filtered
 127 image therefore serves as an estimate of the local back-
 128 ground intensity. Subtracting the large-kernel median
 129 image from the original slice yields an intermediate im-
 130 age in which small- and medium-scale fluorescent fea-
 131 tures are emphasized relative to their local background.
 132 However, this operation also amplifies single-pixel noise
 133 artifacts and hot pixels, which exhibit high local con-
 134 trast and are not suppressed by the background sub-
 135 traction alone. To mitigate this effect, we compute a
 136 second residual by subtracting the small-kernel (3×3)
 137 median-filtered image from the original slice, producing
 138 an estimate of high-frequency noise. Finally, subtracting
 139 this noise estimate from the background-subtracted im-
 140 age suppresses isolated hot pixels while preserving con-
 141 tinuous medium-scale fluorescent structures[fig.9]. The
 142 resulting image preferentially highlights features of in-
 143 terest, such as capillaries and nerve fibers, while reduc-
 144 ing contributions from both large-scale background vari-
 145 ations and pixel-level noise.

146
 3.4. *Postprocessing*
 147 Following preprocessing and segmentation, both
 148 pipelines yield a volumetric stack of binary images rep-

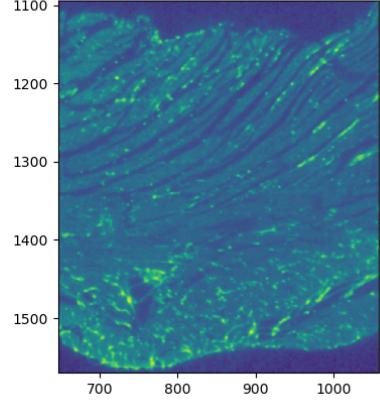


Figure 7. The small median image of the same slice (3×3)

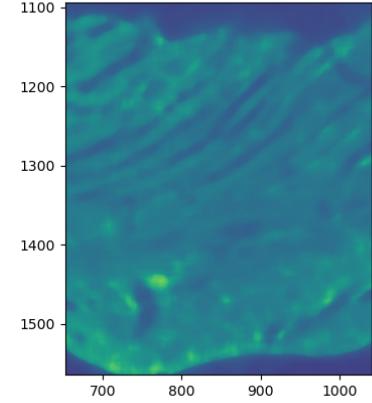


Figure 8. The large median image of the same slice (15×15)

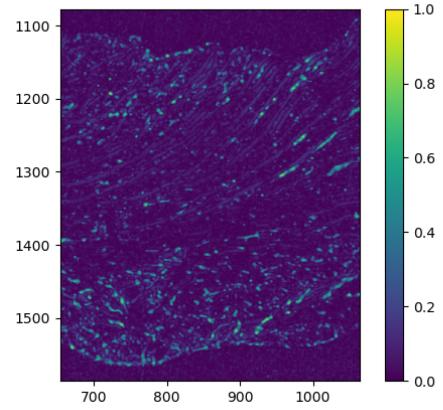


Figure 9. The subtracted I_3 and I_{15}

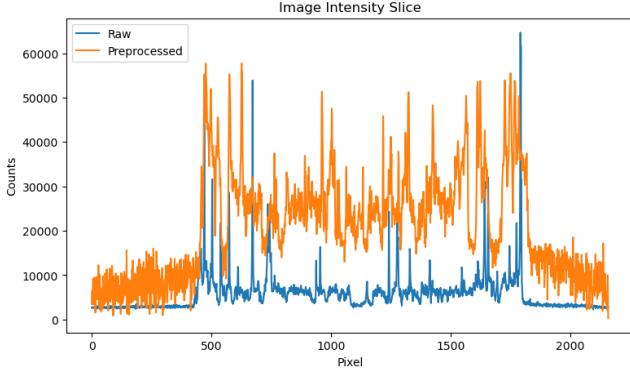


Figure 10. The raw and pre-processed intensities of a 1d slice of data through the x axis

resenting the structures of interest. Here, to take advantage of the well-defined spatial format and to clean up the data, we apply a kernelized erosion/dilation filter. First, to connect structures over the sparser z axis, we used a 1x1x5 kernel in dilation then erosion for 5 iterations. Then, to connect the shapes across the xy axis, we use a standard 3x3x3 kernel dilation/erosion loop once. To enable efficient visualization and downstream analysis, these binary volumes are converted into an optimized mesh representation with the marching cubes algorithm Lorensen & Cline (1987). The resulting meshes can be rendered efficiently on standard hardware and provide a compact representation of the original segmented volume. Finally, to reduce footprint even further and remove residual noise, we apply a number of faces filter, where any floating shape with fewer than 20 faces was removed.

4. RESULTS

4.1. Preprocessing Results

Applying intensity normalization and the asinh transform produced slices with consistent intensity profiles, reducing z-axis variation across the stack. Subsequent CLAHE effectively enhanced local contrast, making small fluorescent structures more distinct against background tissue.

Qualitatively, structures such as capillaries and nerve fibers were more visible post-processing, providing a more uniform input for segmentation. Quantitative intensity metrics indicated a reduction in inter-slice variance[fig.11]. The detrended coefficient of variation decreased from 0.42 to 0.28 over the average intensities of the images, demonstrating improved intensity homogeneity.

4.2. U-Net Feature Detection

The 9-layer U-Net model demonstrated stable convergence over the full 50-epoch training process, achieving a

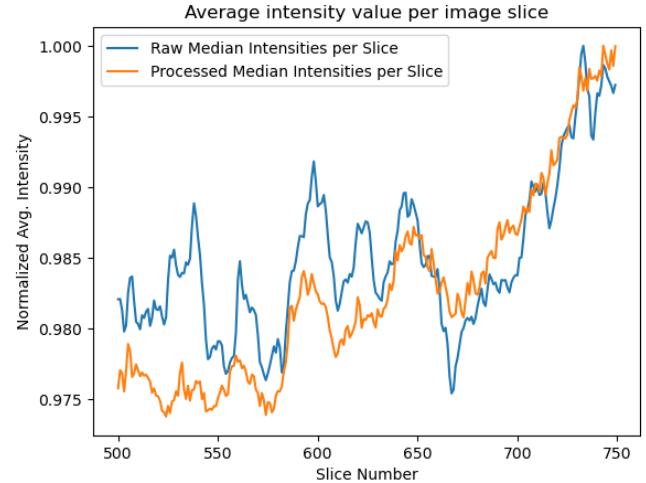


Figure 11. Average intensity profiles across a subset of slices before and after preprocessing. Preprocessing reduces inter-slice variability while preserving large-scale structural features.

Tversky-Focal Loss of 0.55 on the validation set. While the model achieved a high pixel-wise accuracy of 0.98, the Dice Coefficient of 0.29 (Dice Loss of 0.71) highlights the challenge of segmenting fine, sparse structures. This discrepancy is likely due to the extreme class imbalance, where the background dominates the loss calculation despite the use of Tversky-Focal weighting. Segmentation performance was strong for larger, contiguous structures, such as major blood vessels, which were consistently identified with well-defined boundaries across adjacent slices. However, smaller structures like capillaries, which are represented by small or even single-pixel values, were not well segmented. This limitation may be due to the loss function disproportionately prioritizing larger structures (comprising thousands of pixels) over smaller, sparse features. To address this, future work may introduce an additional class for capillaries, or focus on expanding the training dataset, as typical U-Net implementations for similar biological imaging tasks often utilize training sets of over 500 annotated images. In our study, we evaluated the method with a smaller set of 32 images, which likely contributed to the model's difficulty in segmenting fine structures. To partially mitigate this issue, we used the union of U-Net predictions with Difference of Medians masks to improve the segmentation of blood vessels.

4.3. Difference of Medians

The Difference of Medians approach worked well to remove noise and background features. Applying the filter decreased average image intensity from 11890 to 1054, indicating a decrease in the image noise floor and

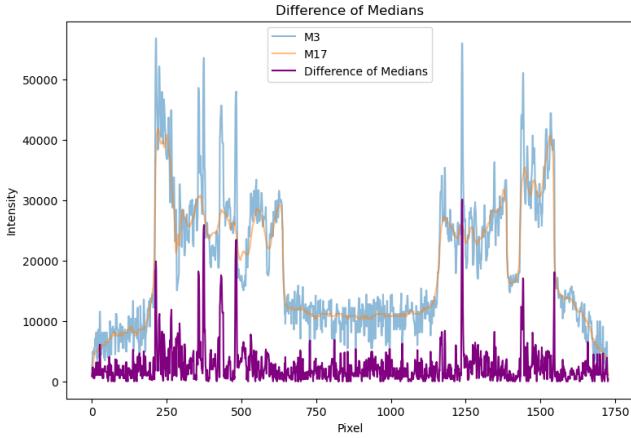


Figure 12. A one dimensional slice illustrating the medium-scale feature detection by the Difference of Medians approach.

216 background structures. The average signal pixel value
 217 also went from 45672 to 16933, making a signal to noise
 218 increase from 3.8 to 16.0. Further, the signal peaks were
 219 both more well-defined and simpler to pick out with an
 220 intensity threshold. Though medium features were de-
 221 tected well, this method scales poorly to larger feature
 222 size band passes. With a time complexity of roughly
 223 $O(n \cdot m \cdot I^2 \log(I))$ Where I is the size of the large me-
 224 dian kernel, this method becomes infeasible for large
 225 features. Additionally, the median kernel will become
 226 less effective at selecting larger features as the feature
 227 contributes more pixels to the median kernel.

228 5. APPLICATION AND FUTURE WORKS

229 The final models produced by this pipeline enable ef-
 230 ficient visualization and analysis of the biological struc-
 231 tures on standard computational hardware. By convert-
 232 ing the segmented volumes into optimized mesh rep-
 233 resentations, we significantly reduce the computational
 234 footprint, allowing the models to be rendered and ex-
 235 plored on a variety of platforms. Both models fit within
 236 a single game file of under 1 GB, demonstrating the fea-
 237 sibility of running such large-scale biological datasets on
 238 modest systems. Figure 13 shows a connectivity analy-
 239 sis that is simple to perform with data in this format,
 240 showing potential utilities that would be impossible with
 241 the original dataset.

242 Looking ahead, there are several avenues for further
 243 development and refinement. One simple improvement
 244 could be the implementation of a blind point spread de-
 245 convolution. Algorithms such as the Richardson Lucy
 246 transform have been modified to both derive a pseudo-
 247 psf and operate with a varying psf across an image.
 248 The U-Net model could include separate segmentation
 249 classes of larger vessels and smaller capillaries within the

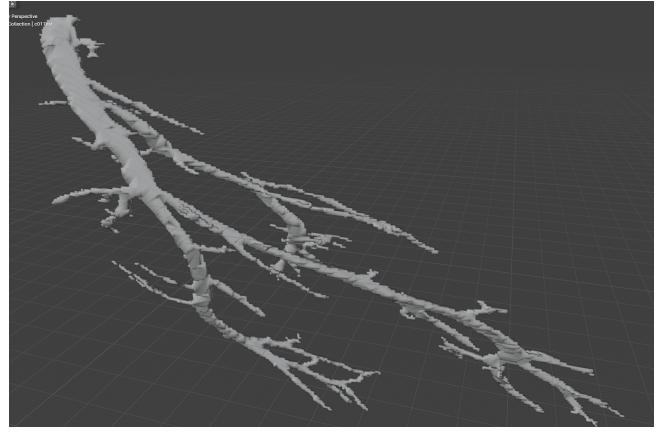


Figure 13. Connectivity analysis within the game engine. By selecting all topologically connected faces, a single vascular ‘tree’ can be isolated from the heart wall. This demon-
 strates the pipeline’s ability to preserve structural continuity, enabling future pathfinding and transport modeling.

250 model. Given the challenges faced in segmenting smaller
 251 structures, a separate class for small features could re-
 252 duce the impact of lopsided data. Additionally, increas-
 253 ing the size of the training dataset could improve model
 254 performance. Given that current U-Net implemen-
 255 tations for similar biological imaging tasks often rely on
 256 training sets with over 500 annotated images, a larger
 257 training dataset would likely yield better generalization
 258 and segmentation of small-scale features. There is also
 259 significant work on 3D U-Net implementations, which
 260 could leverage the z continuity of these structures for
 261 improved performance at the cost of a more complicated
 262 training data preparation and a higher memory cost in
 263 training.

264 Furthermore, while the current dilation/erosion
 265 pipeline for postprocessing is effective, it remains a rel-
 266 atively simplistic method for bridging voxelized struc-
 267 tures. There is significant room for improvement in this
 268 area, as more sophisticated algorithms for voxel con-
 269 nectivity and structure bridging exist in graph-based or
 270 morphological techniques. These algorithms are being
 271 applied on similar datasets like neuron tracing to help
 272 improve the continuity and accuracy of segmented struc-
 273 tures, particularly in regions where adjacent structures
 274 are sparsely connected across slices.

275 In summary, while the presented pipeline provides a
 276 functional and efficient means of segmenting and visual-
 277 izing large-scale biological datasets on modest computa-
 278 tional resources, further optimization and refinement of
 279 segmentation models, training sets, and postprocessing
 280 techniques will be necessary to address the challenges of
 281 segmenting smaller, more intricate structures and fur-

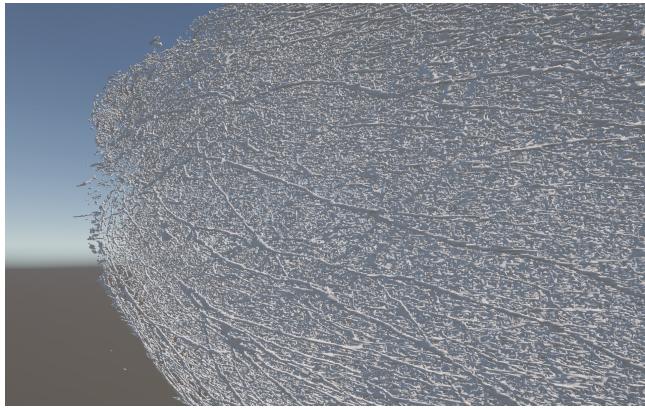


Figure 14. A screenshot of the nerve structure as it appears in the game engine.



Figure 15. An image from within the heart of the blood vessel channel.

ther enhance the utility of this method for biological research.

6. DATA AND CODE

- GitHub for Data Pipeline - <https://github.com/Thomaslund1/Heart-Scan-Pipeline.git>
- GitHub for Demo Render Shown - <https://github.com/Thomaslund1/Heart-Scan-Unity-Demo.git>
- Home of the Human Protein Atlas - <https://www.proteinatlas.org/>

REFERENCES

291 Lorenzen, W. E., & Cline, H. E. 1987, ACM SIGGRAPH 298
 292 Computer Graphics, 21, 163, doi: [10.1145/37402.37422](https://doi.org/10.1145/37402.37422) 299

293 Ronneberger, O., Fischer, P., & Brox, T. 2015, in Medical 300
 294 Image Computing and Computer-Assisted Intervention 301
 295 MICCAI 2015, ed. N. Navab, J. Hornegger, W. M. Wells 302
 296 & A. F. Frangi (Cham: Springer International 303
 297 Publishing), 234–241 304
 298 305

Uhlén, M., Fagerberg, L., Hallström, B. M., et al. 2015, Science (New York, N.Y.), 347, 1260419, doi: [10.1126/science.1260419](https://doi.org/10.1126/science.1260419)

Zekri, M. A., & Lang, I. 2024, Methods in molecular biology (Clifton, N.J.), 2772, 353, doi: [10.1007/978-1-0716-3710-4_27](https://doi.org/10.1007/978-1-0716-3710-4_27)

Zuiderveld, K. 1994, in Graphics Gems IV (USA: Academic Press Professional, Inc.), 474–485